

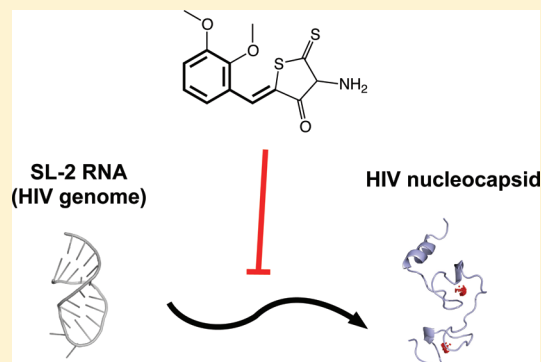
Identification of HIV-1 Inhibitors Targeting the Nucleocapsid Protein

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S Supporting Information

ABSTRACT: The HIV-1 nucleocapsid (NC) is a RNA/DNA binding protein encoded within the Gag polyprotein, which is critical for the selection and chaperoning of viral genomic RNA during virion assembly. RNA/DNA binding occurs through a highly conserved zinc-knuckle motif present in NC. Given the necessity of NC–viral RNA/DNA interaction for viral replication, identification of compounds that disrupt the NC–RNA/DNA interaction may have value as an antiviral strategy. To identify small molecules that disrupt NC–viral RNA/DNA binding, a high-throughput fluorescence polarization assay was developed and a library of 14 400 diverse, druglike compounds was screened. Compounds that disrupted NC binding to a fluorescence-labeled DNA tracer were next evaluated by differential scanning fluorimetry to identify compounds that must bind to NC or Gag to impart their effects. Two compounds were identified that inhibited NC–DNA interaction, specifically bound NC with nanomolar affinity, and showed modest anti-HIV-1 activity in ex vivo cell assays.



INTRODUCTION

The HIV-1 nucleocapsid (NC) protein is a small basic 7 kDa protein encoded within the Gag polyprotein of the infected cell and is cleaved from Gag by the viral protease during maturation (for review, please see Muriaux and Darlix¹). NC provides viral genomic binding specificity allowing viral RNA selection from among the cellular host RNAs, as well as the annealing of the tRNA^{lys} to the primer-binding site.^{2–5} The NC protein is also required for proper dimer formation of genomic RNA in the virus nucleocapsid and in later stages is critical for facilitating reverse transcription.^{6,7} Moreover, it has been shown that altered NC processing from the precursor Gag polyprotein by the viral protease during maturation affects stability of the RNA dimer and thus impairs infectivity of the virus.⁸

NC interacts with a 120 nt RNA segment via hydrophobic and hydrophilic interactions, termed the ψ -site, which is located between the viral 5' long terminal repeat (LTR) and the *gag* start codon in the genomic RNA.⁹ The structure of NC bound to stem-loop 3 (SL-3), one of four stem-loops present in the ψ -site, was solved and shown to be composed of two highly conserved CCHC-type zinc knuckles (a Cys-Xaa₂-Cys-Xaa₄-His-Xaa₂-Cys motif) that directly interact with RNA/DNA.^{10,11} Although the NC protein binds RNA with high affinity, the NC protein has been shown to bind a broad variety of nucleotide sequences and synthetic oligonucleotides.¹

Given the required role of NC in genomic RNA selection and interaction during viral encapsidation, as well as early and late postentry events, it is considered a potential antiviral target. Moreover, the highly conserved NC zinc-knuckle motif, found in all HIV-1 subtypes, is required for viral RNA interaction and presumably has a low tolerance for drug-resistance muta-

tions.^{1,12} These biological properties of NC make it ideal for small molecule targeting. Screening efforts to identify compounds that directly disrupted NC–RNA interaction were undertaken in the early 1990s, resulting in inhibitors that promoted zinc ejection. Although these identified compounds demonstrated antiviral effects, they had a number of shortcomings, including NC specificity and cellular toxicity.^{12–15} Newer reported compounds that disrupt NC–RNA/DNA interaction include Trp-containing peptides, nucleomimetics, RNA aptamers, and gallein-related compounds.^{16–21} Interference of the NC–RNA/DNA binding mechanism by targeting the SL-3 stem-loop, rather than the NC protein, has also led to inhibitors that have not yet been tested in antiviral assays.^{12,22} Recently, Shvadhak and colleagues have focused on the inherent NC chaperone activity for RNA and have identified five compounds that disrupt NC–cTAR DNA interaction, a stem-loop sequence complementary to the transactivation response element, without coordinating through the zinc-knuckle/finger motifs.²³ However, these compounds have yet to be evaluated for anti-HIV-1 activity in cellular assays.²³ Chemotypes of N-substituted S-acyl-2-mercaptobenzamides (SAMTs) have been identified that disrupt NC zinc coordination, resulting in NC protein aggregation and diminished protease processing.^{24–26} Thus, NC inhibitors can have a number of distinct modes of action.^{12,25} Despite the significant screening efforts for identification of NC inhibitors, the lack of compound specificity as well as cellular toxicity of

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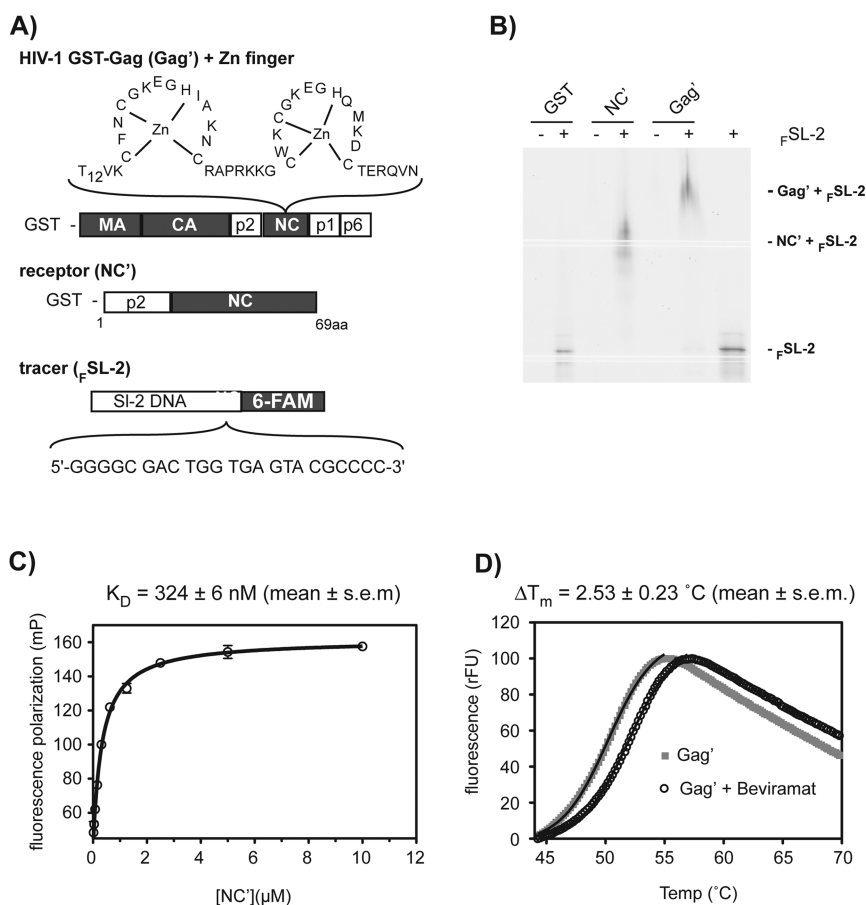


Figure 1. Methodology for identifying compounds that disrupt nucleocapsid–RNA/DNA interaction. (A) Fluorescence polarization (FP) assay components. Shown are the N-terminal GST-tagged Gag (Gag') fusion protein and N-terminal GST-tagged receptor nucleocapsid (NC') fusion protein, containing the spacer peptide 1 (p2) and nucleocapsid domains. NC and Gag are from HIV-1_{NL4-3}. NC contains two zinc-knuckle motifs that interact with viral DNA or RNA. The 3'-6-carboxyfluorescein-labeled SL-2 DNA (pSL-2) is a tracer molecule that interacts with the NC' zinc-knuckle motifs. (B) Electrophoresis mobility shift assay confirms interaction of pSL-2 with NC' or full-length Gag'. Note that GST alone does not shift pSL-2. (C) A fluorescence polarization binding study of pSL-2 and NC' interaction revealed a K_D of 324 ± 6 nM (mean \pm SEM). The data shown represent the mean \pm SEM of three independent experiments performed in duplicate. (D) Thermal protein denaturation of Gag' or maturation inhibitor bevirimat–Gag' complex using differential scanning fluorimetry (DSF). The maturation inhibitor bevirimat increased the thermal stability of the Gag' protein. The black line indicates a nonlinear curve fit based on the Boltzmann equation. The result shown is one representative measurement from three independent experiments.

compounds identified has hampered the progression of NC antivirals to the clinic.

To identify compounds that disrupt NC–RNA/DNA interaction, we developed a high-throughput screening (HTS) platform utilizing fluorescence polarization (FP) as a primary screen and differential scanning fluorimetry (DSF) as a secondary screen. The combination and order of the assays allowed us to first identify compounds that disrupted NC–DNA interaction and then in the secondary screen determine which of the compounds required NC binding for the disruption effect. A total of 14 400 druglike compounds were systematically screened. Five compounds were selected based on their NC–DNA disruption and NC binding affinity profiles. Of the five compounds selected, two demonstrated modest anti-HIV-1 activity in tissue culture infection assays. These compounds may provide a starting point for developing molecules with potential anti-HIV-1 effects.

RESULTS

Establishment and Validation of Screens for Compounds That Disrupt Nucleocapsid–DNA Interaction. A

fluorescence polarization (FP)-based competitive equilibrium-binding assay was utilized as the primary, high-throughput screen (HTS) to identify small molecules (hits) from the HitFinder library that disrupt HIV-1 nucleocapsid (NC) protein–DNA interaction. The small molecule library collection from Maybridge contains 14 400 selected compounds with druglike diversity, which complies with Lipinski's rule of five. The FP-based HTS employed a simple one-step "mix and measure" FP design, which was ideal for miniaturization and HTS automation. A 6-carboxyfluorescein-labeled SL-2 DNA (pSL-2), termed the tracer, was developed to interact with the zinc-knuckle motifs present in a GST-p2-NC fusion protein (NC'), termed the receptor (Figure 1A, B and Supporting Information Figure 1A, B). The DNA bases selected for pSL-2 correspond to the 12 RNA bases of the viral SL-2 RNA, which provide for contact with the NC zinc-knuckle motifs.²⁷ Given that several studies have suggested that the spacer peptide 1 (SP1 or p2) influences RNA packaging and dimerization, the spacer was included in the GST-p2-NC fusion protein reporter.^{28–30}

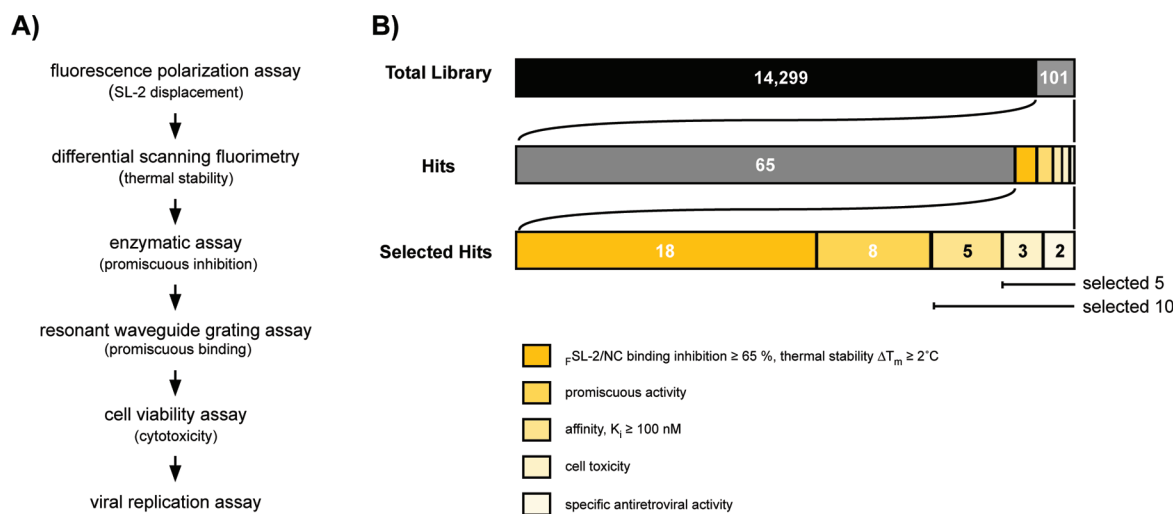


Figure 2. Overview of the strategy for identifying compounds that disrupt HIV-1 nucleocapsid/RNA interaction. (A) Outline of the experimental procedure. (B) Enumeration of hits during library screening. By use of a fluorescence polarization assay to detect small molecule displacement of 3'-6-carboxyfluorescein-labeled SL-2 DNA (FSL-2) from binding to the nucleocapsid (NC') protein, 14 400 compounds were screened and 101 compounds (hits) were identified as displacing FSL-2 . The 101 hits were retested for NC'- FSL-2 displacement, and 65 did not show activity or demonstrated $<60\%$ FSL-2 displacement activity and were eliminated from further study. Of the remaining 36 compounds (selected hits), 18 failed to show a significant shift in thermal stability when the compound-Gag complexes were evaluated by differential scanning fluorimetry assay, 8 displayed promiscuous β -galactosidase inhibition, and 5 were excluded because of low affinity binding to NC'. Of the five remaining selected hits, three hits showed cellular toxicity at higher compound concentrations, whereas two compounds demonstrated both low cellular toxicity and inhibition of HIV-1 replication in CD4⁺ T cells.

SL-2 binding to NC' and the Gag polyprotein (Gag') was first validated with an electrophoresis mobility shift assay (EMSA) (Figure 1B), as well by native PAGE utilizing a fluorescent protein fusion variation of NC' (Supporting Information Figure 1C). Importantly, FSL-2 binding to NC' had a stabilizing effect resulting in a homogeneous protein form rather than an oligomerized form, an often-reported problem when using the disordered apo-form of zinc-knuckle/finger proteins (Figure 1B and Supporting Information Figure 1C).¹ The robust interaction of FSL-2 with NC' revealed an 8-fold change of fluorescence polarization which provided an excellent signal-to-noise (S/N) ratio, high to low (H/L) signal ratio, and a Z' factor of 0.84 ± 0.02 (mean \pm sd) (Supporting Information Figure 1D,E).³¹ The NC'- FSL-2 dissociation constant was determined to be $324 \pm 6\text{ nM}$ (Figure 1C).

To ensure that we identified compounds that bound to NC in the context of full-length Gag to disrupt FSL-2 interaction, thereby potentially increasing in situ specificity, a differential scanning fluorimetry (DSF) screen was established. The rationale for the inclusion of this screen was to allow a rapid throughput and determination if binding of the compound to NC increased or decreased the melting temperature, thus indicating that the Gag-compound complex was more thermally stable than Gag alone. The feasibility of this methodology was validated by denaturing the Gag' protein in the presence and absence of the HIV-1 maturation inhibitor bevirimat (PA-457), which is reported to function in blocking maturation via Gag interaction.³² As seen in Figure 1D, Gag' in complex with bevirimat has a higher melting temperature (T_m) than Gag' alone, consistent with the notion that bevirimat interacts with Gag and increases its thermal stability. This finding supports the use of the DSF assay for identifying compounds (selected hits) that bind to and increase or decrease NC' and Gag' stability. To place the screening and hit validation strategy in a temporal perspective, a flowchart is

shown in Figure 2A, with Figure 2B providing an enumeration of the successful hits identified during screening and the cellular validation process.

Compounds That Disrupt Nucleocapsid-DNA Interaction. Use of the FP HTS for interrogating the 14 400-compound HitFinder library, resulted in 101 hits that disrupted NC'- FSL-2 interaction (Figure 2B). Hit identification was based on two quantitative criteria: first, disruption of NC'- FSL-2 interaction was required to be $>50\%$ at $10\ \mu\text{M}$, and second, intrinsic compound fluorescence was negated by comparing FP to total fluorescence (Supporting Information Figure 1D).³³ Next, we determined whether the 101 hits identified were functioning to disrupt NC'- FSL-2 by binding to NC' in the context of the polyprotein Gag and forming a stable complex. Of the 101 compounds, 36 displayed $\Delta T_m > 2^\circ\text{C}$ when analyzed by DSF, the criterion established for selection, but only 18 of the 36 compounds also demonstrated inhibition of NC'- FSL-2 interaction by more than 60% (Supporting Information Figure 2A). These 18 compounds were selected for further analyses (Figure 2).

To determine if the 18 selected hits were promiscuous inhibitors, two enzymes, β -galactosidase and chymotrypsin commonly used to uncover compounds that bind promiscuously and inhibit enzymatic function, were used for evaluation.³⁴ Of the 18 selected hits, 8 were eliminated, since they demonstrated $>10\%$ inhibition of these enzymes at $10\ \mu\text{M}$ (Supporting Information Figure 2B). The remaining 10 compounds were selected for further evaluation (Figure 2).

Biochemical Profiling: Affinity and Specificity Evaluation of the 10 Selected Compounds. In order to quantify the inhibitor-activity relationship of the 10 compounds, the K_i values were determined using the FP assay (see the Experimental Section for details).³⁵ Newly synthesized compounds from Maybridge (see the Experimental Section for details) were utilized to avoid storage-associated effects. All

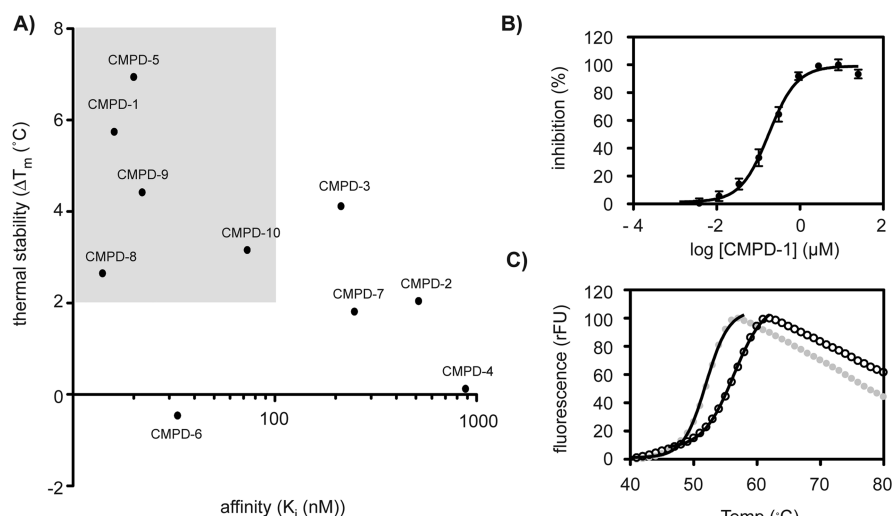


Figure 3. Biochemical characterization of 10 compounds selected by fluorescence polarization and differential scanning fluorometry assays. (A) Correlation of DSF-based thermal stability versus FP-based K_i values of resynthesized compounds. Only compounds with $\Delta T_m \geq 2$ °C and $K_i \leq 100$ nM were selected for further evaluation. (B) Binding curve of compound 1 with NC' in FP-based displacement experiment reveals a high affinity ($K_i = 18 \pm 14$ nM). Shown is the mean \pm sd of a representative experiment from three experiments. (C) DSF-based stability analysis of the 1–Gag' complex. Compound 1 enhances the stability of Gag' by $\Delta T_m = 5.7 \pm 1.0$ °C (gray circles = control; black open circles = Gag' + 1; nonlinear fit = black line).

10 compounds were found to have K_i values in the nanomolar range (Figure 3A,B). Moreover, the order of addition of compounds, NC', or $_F$ SL-2 did not alter compound effects or K_i (data not shown). To determine potential promiscuous activity, the 10 selected compounds were evaluated at higher concentrations, utilizing DSF as well as the galactosidase and chymotrypsin enzymatic activity assays (data not shown). Five of the 10 compounds, 2, 3, 4, 6, and 7, displayed weaker affinity ($K_i \geq 100$ nM), thermal stability ($\Delta T_m \leq 2$ °C), and nonspecific effects at higher concentrations (Figure 3), thus eliminating these compounds from further studies.

The remaining five compounds, 1, 5, 8, 9, and 10, were next evaluated by EMSA for their ability to disrupt the NC'– $_F$ SL-2 complex. The preformed NC'– $_F$ SL-2 complex was incubated with the compounds, separated by native PAGE, and the $_F$ SL-2 intensity was evaluated by fluorescence scanning. All compounds displaced $_F$ SL-2, confirming their disruption of the NC'– $_F$ SL-2 interaction (Figure 4A). Furthermore, studies indicated that the order of NC', $_F$ SL-2, or compounds to the reaction mixture did alter the capacity of any of the five compounds to disrupt the NC'– $_F$ SL-2 complex (data not shown). This suggests that the compounds do not require prebinding to NC' to impart their SL-2 inhibitory affect.

To determine NC'–compound stoichiometry, all five compounds were evaluated by a resonant waveguide-grating-based assay employing SRU BIND technology.³⁶ Resonant waveguide-grating-based assessment has been useful for the evaluation of protein–drug interaction, protein–DNA inhibitors, as well as the identification of promiscuous binders to proteins.³⁶ To evaluate the five compounds, we immobilized NC' on an optical biosensor, then monitored compound binding by measuring the peak wavelength value shift over time. The plateauing of NC'–compound binding kinetics implies a specific binding mode for compounds 1, 5, 8, and 10, with the NC'–compound stoichiometry determined to be approximately 1:1 (Figure 4B). Although compound 9 displayed a plateauing of NC'–compound binding kinetics, the stoichiometry was determined to be approximately 1:2.

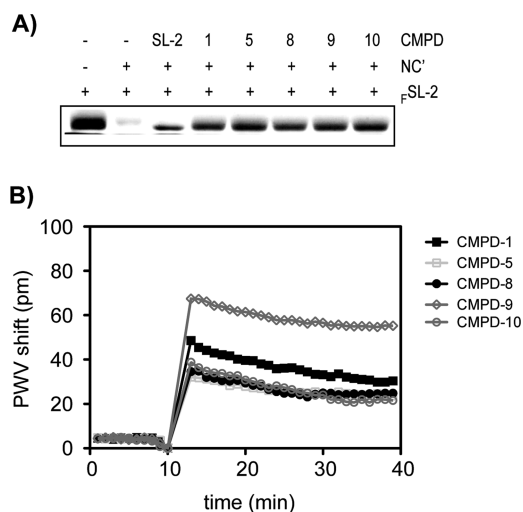
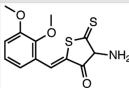
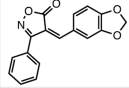
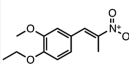
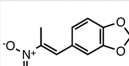
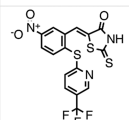


Figure 4. Biochemical characterization of five selected compounds. (A) EMSA-based analyses of the binding of 1, 5, 8, 9, or 10 to NC' showing the displacement of $_F$ SL-2 from the NC'– $_F$ SL-2 complex or the internal control utilizing 10 μ M unlabeled SL-2 DNA. (B) Resonant waveguide-grating-based analyses of the binding of 1, 5, 8, 9, or 10 to immobilized NC'. Real-time kinetics of compound binding was monitored by peak wavelength value shifts over time.

This finding is consistent with two molecules of compound 9 binding to NC. In summary, these findings indicate that the five selected compounds had $K_i \leq 100$ nM, disrupted $_F$ SL-2 binding to NC', and recognized the NC domain in the Gag polyprotein and that four of the five compounds demonstrated 1:1 stoichiometry for NC'–compound interaction. Table 1 summarizes the chemical and biochemical findings for the five compounds.

Biological Profiling: Cellular Toxicity and Anti-HIV-1 Activity of Compounds 1, 5, 8, 9, and 10. The cellular toxicity of compounds 1, 5, 8, 9, and 10 were evaluated at 0.1 and 10 μ M at 3 and 7 days after addition to SupT1 T cells, a CD4⁺ T cell line (Figure 5A,B). At 10 μ M, compounds 5, 8, 9,

Table 1. Summary of the Biochemical and Biological Characterization of Compounds 1, 5, 8, 9, and 10^a

Sample ID	MB ID	Name	PubChem ID	MW (KDa)	K _i (FP) nM (mean ± s.e.m.)	DSF (°C) Δ T _m	optical biosensor (SRU bind)	% inhibition (p24 assay) at 200 nM
CMPD-1		(Z)-4-amino-2-(2,3-dimethoxybenzylidene)-5-thioxodihydrothiophen-3(2H)-one	26532231	295.38	18 ± 14	4.8 ± 0.4	n = 1:1; np ^a	43 ± 8
CMPD-5		(Z)-4-(benzo[d][1,3]dioxol-5-ylmethylene)-3-phenylisoxazol-5(4H)-one	26533388	293.27	20 ± 5	6.3 ± 0.1	n = 1:1; np	33 ± 10
CMPD-8		(E)-1-ethoxy-2-methoxy-4-(2-nitroprop-1-en-1-yl)benzene	26541579	237.25	14 ± 7	2.2 ± 0.6	n = 1:1; np	50 ± 12
CMPD-9		(E)-5-(2-nitroprop-1-en-1-yl)benzo[d][1,3]dioxole	26529289	207.18	22 ± 9	3.4 ± 0.7	n = 1:1.7; np	18 ± 16
CMPD-10		(Z)-5-(5-nitro-2-((5-(trifluoromethyl)pyridin-2-yl)thio)benzylidene)-2-thioxothiazolidin-4-one	26540561	443.44	73 ± 50	2.4 ± 0.6	n = 1:1; np	0 ± 0

^anp = nonpromiscuous binder.

and **10** exhibited >90% cytotoxicity at 7 days (Figure 5A). At 30 μM compound **1** demonstrated >70% toxicity within 6 days (not shown). At the lower concentration of 0.1 μM, **1**, **5**, **8**, **9**, and **10** demonstrated 0–38% cell toxicity within 7 days (Figure 5A), whereas at 1 μM, **5**, **9**, and **10** compound demonstrated >70% cell toxicity (not shown). In contrast to compounds **5**, **9**, and **10**, compounds **1** and **8** evaluated at 0.1 or 1 μM exhibited <3% cell toxicity within 7 days (Figure 5A). Similar cellular toxicity patterns were seen with the HeLa cell line, as well as with activated, primary CD4⁺ T cells (not shown).

We next evaluated anti-HIV-1 activity of the five selected compounds on activated, primary CD4⁺ T cells. A concentration of 0.2 μM compounds was evaluated based on compound–NC' affinity (Figure 3A) and the range of cellular toxicity (Figure 5A). Activated, primary CD4⁺ T cells were cultured for 3 days in the presence of compounds **1**, **5**, **8**, **9**, and **10** and then infected with 100 ng of p24 of HIV_{LAI}. After 9 days the level of HIV-1 infection in CD4⁺ T cells was enumerated through detection of infected, intracellular p24 positive T cells present in the culture (Figure 5B). Relative to the control cultures, compound **10** did not exhibit a viral inhibitory affect. In contrast, compounds **1**, **5**, **8**, and **9** modestly reduced T cell infections from 18% to 50%, with compounds **1** (43 ± 8% viral inhibition), **5** (33 ± 10% viral inhibition), and **8** (50 ± 12% viral inhibition) showing the greatest anti-HIV affect (Figure 5B and Table 1).

Compounds **1** and **8** were selected for further anti-HIV-1 evaluation given their slightly better antiviral activity than compounds **5**, **9**, and **10**. Moreover, the toxicity of **5**, **9**, and **10** precluded additional cell-based studies. The viral spreading assay used for the studies shown in Figure 5B allows all steps of viral replication to be targeted by an inhibitory compound. To determine inhibitory EC₅₀ values for compounds **1** and **8**, we utilized a HIV DNA transient transfection assay to produce virus from HeLa cells in the presence or absence of each compound. This procedure bypasses the early stages of the viral replication cycle and provides information on compound activity in later stages of viral replication (ie, after integration). Viral production was quantified by determining p24 amounts.

When this assay was utilized, the EC₅₀ for compound **1** was determined to be 3.5 μM, whereas for compound **8** the EC₅₀ was 0.32 μM (Figure 5C).

DISCUSSION

The primary role of NC during viral replication and its inability to tolerate mutations make it an ideal target for small molecule interaction to disrupt HIV-1 replication.³⁷ Moreover, a 20–30% reduction in Gag processing results in a 3 log decrease in viral production.³⁸ We have identified two compounds that interfered with NC–DNA and Gag–DNA interactions in biochemical assays and viral replication in primary CD4⁺ T cells. A simple series of assays were developed that required compounds to disrupt NC–DNA interaction for positive selection and a secondary screen to identify compounds that bound to NC' to impart their disruptive affect. This second screen ensured that the mode of action of the compound requires NC' binding and is not simply the result of compound interacting with DNA. Moreover, the use of the Gag polyprotein for later compound assessment takes into account the knowledge that NC domain and viral RNA interaction takes place in the context of the Gag polyprotein in host cells. The use of the 14 400 druglike, small molecule HitFinder library for screening may have increased the probability for identifying compounds with favorable biological and chemical activities, as well as possibly reducing cellular toxicity and promiscuous inhibitory activities. Compounds **1**, **5**, **8**, **9**, and **10** formed stable complexes with NC' (Figure 3A and Table 1) and, except for compound **9**, demonstrated NC–compound binding stoichiometries of 1:1 (Table 1).

Of the five compounds selected for biological evaluation, compounds **1** and **8**, which demonstrated modest anti-HIV affect in primary CD4⁺ T cells at 200 nM (**1**, 43 ± 8% viral inhibition; **8**, 50 ± 12% viral inhibition), had the lowest FP K_i values, with no overt promiscuous activity (Figure 5 and Table 1). The inhibitory evaluation of compounds **1** and **8**, the least toxic of the five compounds, in the viral packaging assay yielded EC₅₀ of 3.5 μM for compound **1** and 32 nM for compound **8**. Interestingly, although compound **8** showed comparable

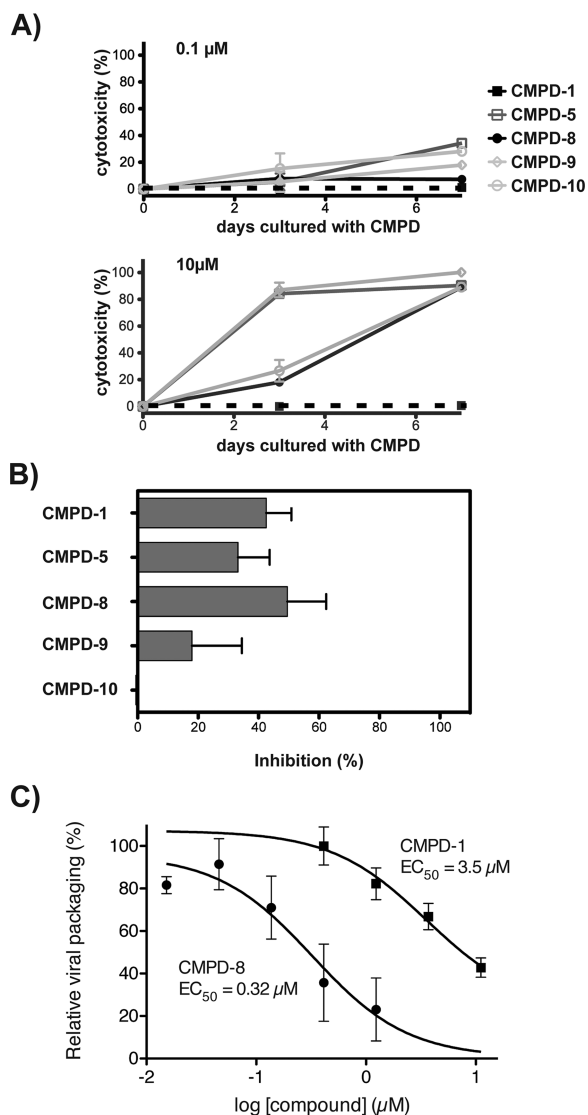


Figure 5. Cellular toxicity and anti-HIV-1 evaluation of five selected compounds. (A) Cellular toxicity evaluation of 1, 5, 8, 9, or 10. Cellular viability over time of SupT1 T-cells was evaluated in the presence of 0.1 or 10 μM each of the five compounds using a fluorescence-based cytotoxicity assay. No compound control cultures contained 0.2% DMSO, the diluent for the compounds, for comparison. (B) Inhibition of HIV_{LAI} replication in primary CD4⁺ T cells. Cells were cultured with 0.2 μM 1, 5, 8, 9, or 10 for 3 days before exposure to 100 ng of p24 of HIV_{LAI}. Nine days after HIV-1 exposure, cells were evaluated for infection by flow cytometry to determine cellular viral p24, and results were plotted as mean \pm sd of % inhibition. 0.2 μM amprevanir, a protease inhibitor, was used at 5-fold over EC₅₀ as a known inhibitor of viral replication and was found to inhibit viral replication 98% (data not shown). All assays were in triplicate, and shown is one of two replicate experiments. (C) Compounds 1 and 8 disrupt HIV-1 production. The EC₅₀ values of compounds 1 and 8 were determined utilizing a HeLa cell line transiently transfected with plasmid HIV_{NL4-3} DNA in the absence or presence of increasing concentrations of each compound. HeLa cell culture controls contained 1% DMSO (v/v), the diluent used for the compounds. Viral production was determined by measuring p24 production. Results are shown as relative viral packaging in the presence of compounds as compared to DMSO controls. All points are the result of three independent transient transfections, and results are shown as the mean \pm sd of relative viral packaging. EC₅₀ values were determined using nonlinear regression analysis in Prism 5.0d to fit a three-parameter dose–response curve to normalized p24 values.

inhibitory activity in both the viral spreading assay (Figure 5B) and the viral production assay (Figure 5C), compound 1 did not show comparable inhibitory activity in the two assays. Whether the differences in the inhibitory activity of compound 1 reflect targeting of earlier stages of viral replication or cell-based mechanisms supporting viral replication, in addition to the later stages of viral replication, is unclear at the current time and is the subject of investigation. Nonetheless, both compounds exhibited modest antiviral activity.

SMAT-based NCp7 inhibitors have been shown to disrupt protease processing of Gag as the result of compound–NC interaction, followed by inhibitor-mediated NC aggregation resulting in decreased HIV-1 replication in cell culture.²⁵ To evaluate whether the binding of compounds 1 and 8 to NC interfered with Gag polyprotein processing, we monitored p2/NC and NC/p1 cleavage in the presence and absence of both compounds utilizing the cleavage enzyme-cytometric bead array, a Gag-specific cleavage assay (data not shown).³⁹ Our results revealed that compounds 1 and 8 did not interfere with HIV protease enzymatic function or block Gag cleavage. Thus, the inhibitory mechanism of compounds 1 and 8 in cell culture was most likely not the result of either compound interfering with Gag cleavage. Furthermore, and in contrast to the gallein-based inhibitors, compounds 1 and 8 inhibited NC–DNA interaction independent of the order of addition of the compounds or DNA in the FP assay. The gallein-based NC inhibitors require binding to free NC' before the addition of DNA to impart their inhibitory affect.²¹ On the basis of the differing NC binding requirements for gallein-based and compounds 1 and 8 for inhibitory function, our findings might suggest that compounds 1 and 8 bind to different NC locations than the gallein-based inhibitors to impart their anti-HIV effects.

Evident in all compounds obtained are similar chemical motifs, an electron-rich aromatic ring structure in conjugation with an electron-withdrawing nitro substituent or an electron-poor heterocycle (Table 1). A closer evaluation of the five compounds revealed nitroalkenes in compounds 8 and 9, rhodanine in compound 10, and isoxazolone in compound 5, features suggestive of pan assay interference compounds, which can correlate with undesirable chemical properties.⁴⁰ However, the behavior of the five compounds in the resonant waveguide-grating assessment was inconsistent with either reactivity or aggregation (Figure 4B). Moreover, mass spectrometry analysis did not reveal covalent binding between NC' and any of the compounds (Supporting Information Table 1). We next searched the PubChem compound database to determine if screening information was available on any of the five compounds identified.⁴¹ The search indicated that compounds 1, 5, and 8 were not active in seven or more assays against various protein targets, whereas compound 10 were found to be active in one out of seven assays (Supporting Information Table 2). Compound 9 demonstrated promiscuous behavior in some assays, showing activity in 13 of 78 assays, albeit mostly from a related group of anticancer screens. Although the perusal of the PubChem compound database was suggestive of limited protein reactivity, compounds 5, 9, and 10 were cell toxic at moderate compound concentrations. Given the potentially undesirable chemical properties of compounds 5, 9, and 10, as discussed, further assays are required to fully evaluate potential off-target effects.

Interestingly, compounds **1**, **5**, **8**, and **9** have some chemical similarities to E03 and H04, two compounds identified by Shvadchak and colleagues as NC polyprotein binders capable of disrupting DNA binding (Supporting Information Figure 3).²³ However, anti-HIV activities of those two compounds were not evaluated. Both compounds display similar structural features as compounds **8** and **9** regarding the benzene ring structure but have branching oxygen and/or hydroxyl groups in contrast to the pendent nitro groups present on compounds **8** and **9** (Supporting Information Figure 3). In the case of the compounds identified by Shvadchak and colleagues, the oxygen and/or hydroxyl groups likely have a role in NC binding. Moreover, their studies indicated that activity was not coordinated through interaction or zinc ejection but possibly through competition with DNA binding to NC.²³ Given similar chemical structural features of these compounds and the compounds identified in our screen, it is tempting to speculate that our compounds may function to compete with DNA binding to NC.

In conclusion, development and implementation of our screen have resulted in the identification of five compounds that are capable of disrupting p2-NC and Gag polyprotein interaction with SL-2 DNA. Of the five compounds, two were not overtly toxic to primary CD4⁺ T cells and both had modest effects on disrupting HIV-1 spread in tissue culture. Although NC binding is required for the inhibitory effect of the compounds, whether the compounds' activity was due to DNA binding competition resulting from direct inhibition of the zinc knuckle motifs or resulting from disruption of DNA binding caused by allosteric changes due to a distal NC binding site is currently unknown. Efforts are planned to structurally define the NC binding sites of compounds **1** and **8** to provide rational insights for improving new compounds to enhance anti-HIV-1 efficacy. Lastly, given the high-throughput capacity of the FP-based NC' binding assay and the relative ease of use, it will be informative to screen a larger and more diverse chemical library to identify compounds that may have antiviral activity.

EXPERIMENTAL SECTION

Cloning and Recombinant Protein Synthesis. The HIV-1 NL4-3 *p2-nc* gene was cloned into the pGex4T2 vector (GE Healthcare) vector using *Bam*HI and *Xho*I restriction to generate GST-p2-NC (NC'), transformed into BL21DE3(pLysS) (Invitrogen), inoculated into 1–6 L (depending on need) of LB medium with ampicillin (100 µg/mL), and maintained at 28 °C for 16 h. At 0.8 OD_{600nm}, expression was induced with 0.5 mM isopropyl-β-D-thiogalactopyranosid. After 16 h, the cells were harvested by centrifugation (6000g, 15 min) and washed with buffer A (20 mM Tris-HCl, pH 8.0, 500 mM NaCl, 0.5 mM DTT, and 0.5% NP40). Cells were then lysed by the addition of lysozyme and the use of a French press. To avoid bacterial protease activity, a protease inhibitor mix (Complete-Mini protease inhibitor tablets, Roche) and pepstatin A (Sigma-Aldrich) were added. DNA was removed from the lysate by adding 0.04 mg/mL DNase I for 1 h. After separation by centrifugation (30000g, 45 min), the supernatant was loaded onto a GSH column (GE Healthcare) and washed with 20 column volumes of buffer A. The protein was eluted by addition of 20 mM GSH in buffer A and the purity verified by SDS-PAGE. The protein was concentrated using Amicon Ultra-15 filter devices (Millipore). The protein size was confirmed by MALDI MS. All purification steps were conducted at 4 °C. GST-Gag (termed Gag') was produced as reported previously.³⁹

Electrophoretic Mobility Shift Assay (EMSA). 15 µM NC' or Gag' was incubated with 0.5 µM pSL-2 in PBS with 10 nM ZnCl₂. For

the electrophoretic analysis, 15 µL samples were loaded on a NativePAGE Novex Bis-Tris gel (Invitrogen), run, and fluorescence was measured by a Typhoon Trio fluorescence gel scanner (GE Healthcare).

Fluorescence Polarization (FP). The binding reaction was carried out in 384-well, low volume plates (Greiner) using 2.5–10000 nM NC' in PBS, 10 nM ZnCl₂, 0.1% pluronic F127 (Sigma), and 10 nM 3' fluorescein-labeled SL-2 DNA, GGGGCGACTGGTGAG-TACGCCCC (Integrated DNA Technologies). The millipolarization (mP) units were calculated using

$$\text{mP} = \left(\frac{S - PG}{S + PG} \right) \times 100$$

where *S* is the fluorescence of the S-channel, *P* is the fluorescence of the P-channel, and *G* is the instrument factor.

The *K_d* was calculated using the least-squares fit to the following equation:

$$\text{FP} = \text{FP}_{\min} + (\text{FP}_{\max} - \text{FP}_{\min}) \left\{ \frac{(P_0 + L_0 + K_d) - \sqrt{(P_0 + K_d + L_0)^2 - 4(L_0P_0)}}{(2P_0)} \right\}$$

where *P₀* is the total receptor concentration, *L₀* is total ligand concentration, *K_d* is the binding constant, and FP is the fluorescence polarization.

IC₅₀ and *K_i* Determination by FP. The *K_i* was determined by a compound series dilution into 5 µM NC' and 10 nM 6-carboxyfluorescein-labeled SL-2 DNA in PBS, 10 nM ZnCl₂, and 0.1% Pluronic F127. All experiments were in triplicate. The NC' concentration was chosen such that 90% of tracer was bound. Since the concentration of free tracer was not equal to the concentration of total tracer, the Cheng-Prusoff equation could not be used. The IC₅₀ value was calculated using a four-parameter logistic model, and the *K_i* was determined as established by Kenakin 1993:³⁵

$$K_i = \frac{(L_b)(IC_{50})(K_d)}{(L_0)(R_0) + L_b(-R_0 - L_0 + L_b - K_d)}$$

where *R₀* is the total receptor concentration, *L₀* is the total tracer concentration, *L_b* is the bound tracer fraction, and *K_d* is the dissociation constant.

Small-Compound Library and High-Throughput Screening.

The HitFinder library collection from Maybridge contains 14 400 compounds with druglike diversity, and the compounds conform to Lipinski's rule of five with purity of >90% based on mass spectrometry and NMR by the vendor (http://www.maybridge.com/portal/alias_Rainbow/lang_en/tabID_229/DesktopDefault.aspx). For single compounds used for biochemical and biological profiling a purity of >95% was confirmed by HPLC.

The binding reaction was carried out in 384-well low volume plates (Greiner) using 5 µM (tracer bound to receptor, >80%) NC' in PBS, 10 nM ZnCl₂, 0.1% Pluronic F127 (Sigma), and 10 nM of the tracer 6-FAM-labeled SL-2 DNA. The compounds were dissolved in DMSO and used in a final concentration of 10 µM at room temperature. Dispensing and readout were executed using automated liquid handling systems (dispenser Aurora, liquid robot BiomekFX Beckman Coulter) and an EnVision 2104 (PerkinElmer) fluorescence plate reader. NC' and compounds were preincubated at room temperature for 15 min in a volume of 10 µL before 10 nM tracer was added to the solution. The mix was allowed to establish equilibrium, followed by FP readout 15 min later. The plate layout provided for 16 positive controls (tracer without NC') and 16 negative controls (tracer with NC') per plate. The *Z'*-factor was calculated using the equation established by Zhang and colleagues.³¹

Differential Scanning Fluorimetry (DSF). The melting temperatures of protein–compound complexes were determined using differential scanning fluorimetry. An amount of 5 µM Gag' protein was incubated with 5× SYPRO Orange (Invitrogen) and 5 µM compound in 5 mM HEPES, pH 7.5, with 150 mM NaCl. Fluorescence values were monitored in a LightCycler 480 II

(Roche) instrument with temperature increasing at 1 °C/min from 35 to 94 °C. The data were fitted to a Boltzmann sigmoid curve function, and the T_m was obtained as described by Vedadi et al.,⁴² using GraphPad Prism. A shift of 2 °C corresponds to a 4-fold change of the standard deviation and was considered to be a hit.

β -Galactosidase Assay. To determine off-target hit inhibition, the conversion of 0.2 mM ONPG (Sigma) by 0.5 U/mL β -galactosidase (Sigma) was monitored in the presence or absence of 10 μ M selected hit. The reactions were established in PBS with 1% DMSO at 25 °C. The absorption at 420 nm was followed over time using a μ Quant 800 microplate reader (Biotek). The rate was calculated from the initial linear phase of the enzyme reaction, and the normalized inhibition was calculated relative to the rate of the DMSO control reaction.

Chymotrypsin Assay. An amount of 0.5 U/mL chymotrypsin (Sigma) was mixed with 0.2 mM S-2586 (DiaPharma) and 10 μ M selected compounds in 20 mM Tris-HCl, pH 8.3, 200 mM NaCl, 1 mM CaCl₂, and 1% DMSO, all at 25 °C. Absorption at 420 nm was followed over time using a μ Quant 800 microplate reader (Biotek). The initial phase was fitted with a linear regression and the relative rate calculated for each compound.

Resonant Waveguide Grating-Based Assay. To evaluate label-free interaction between NC' and selected compounds, the SRU BIND system (SRU Biosystems) was utilized. An amount of 5 μ M NC' protein was immobilized on the wells of an aldehyde activated biosensor surface 384-well GA3 aldehyde plate (SRU Biosystems). Selected compounds at 25 μ M in PBS with 1% DMSO were added to plated wells. The peak wavelength value (PWV) shift was monitored over time at room temperature in a BIND SCREENER reader (SRU Biosystems). The stoichiometry was calculated according to the vendor's manual using the following equation:

$$PWV_{\text{ligand(expected)}} = \left(\frac{MW_{\text{ligand}}}{MW_{\text{receptor}}} \right) (PWV_{\text{target}})(n)(z)$$

where n is the stoichiometry and z is the target activity in %.

Cellular Toxicity Assay. The SupT1 T cell line or human CD4⁺ T cells were cultured in RPMI1640, containing 10% fetal bovine serum (FBS), 2 mM L-glutamine, 0.1 mM nonessential amino acids (NEAA), 1 mM sodium pyruvate, 100 U/mL penicillin, and 100 μ g/mL streptomycin for all studies (see below for methodology for T cell isolation). Cells were maintained in a 37 °C humidified incubator containing 5% CO₂. Viability of the cells in the presence of selected compounds was measured either by the conversion of the oxidation–reduction indicator resazurin (Alamar Blue) (Invitrogen) to resorufin^{43,44} or by enumerating dead vs viable cells by Trypan Blue dye exclusion by cell counting utilizing a microscope. The 15 000 SupT1 cells were cultured over 7 days with a series of selected compounds (0.01–10 μ M) in 1% DMSO, and the viability was determined in the presence of Alamar Blue according to the manufacturer's protocol. The 585 nm fluorescence value was normalized to a DMSO control. Primary T cells were isolated from blood cells, and in some cases Sup T1 T cells were cultured in the presence of selected compounds (0.01–10 μ M). Viability was determined at days 3 and 6 or 7 by Trypan Blue dye exclusion.

Determination of Compound Anti-HIV-1 Activity. After approval from The Scripps Research Institute (TSRI) Institutional Review Board (documentation is on file), La Jolla, CA, U.S., human blood was obtained from healthy donors from the Normal Blood Donation Center at TSRI. PBMCs were isolated from blood via density gradient centrifugation. Isolation of highly pure CD4⁺ T cells was achieved by depletion using Miltenyi products (Miltenyi Biotek). CD4⁺ T cell purity, >95%, was determined by flow cytometry with appropriate reagents. CD4⁺ T cells were maintained in RPMI1640 medium, supplemented with 10% FBS (HyClone), 100 U/mL IL-2, 2 mM L-glutamine, 0.1 mM NEAA, 1 mM sodium pyruvate, 100 U/mL penicillin, and 100 μ g/mL streptomycin in a 37 °C humidified incubator containing 5% CO₂. After a 1-day stimulation with anti-CD3/CD28 beads (Dynabeads human T-activator CD3/CD28, Invitrogen), 0.2 μ M selected compounds in 1% (w/v) DMSO was

added for 3 days. Then 1×10^5 CD4⁺ T cells were infected with 100 ng of p24 HIV-1_{LAI} virus. Cultures were maintained for 9 days, and infection was determined by p24 flow cytometry analysis. Briefly, HIV-1 infected CD4⁺ T cells were permeabilized and fixed with Cytofix/Cytoperm buffer, followed by addition of PE-conjugated HIV p24 antibody (Beckman Coulter) to a final 1:200 dilution. Data were acquired on Becton Dickinson FACScalibur using CELLQUEST software and analyzed with FlowJo (TreeStar) software. The FDA approved protease inhibitor amprenavir was used as a positive inhibitor of viral replication (data not shown). The significance of the difference between the DMSO control and a test compound was determined using a two-tailed t test.

To determine the effective dose of compound to inhibit viral packaging in HeLa cells, a viral packaging assay was utilized. Selected compounds were added to 2×10^5 HeLa cells in 1 mL of Opti-MEM (Invitrogen) in 12-well plates. After 2 days, cells were transiently transfected with 1 μ g of plasmid DNA of the pNL4-3 infectious molecular clone using Lipofectamine 2000 (Invitrogen) according to the manufacturer's instructions. Cell-free viral supernatants were collected 2 days after transfection and p24 determined by CFAR at the University of California, San Diego, CA, utilizing a commercial p24 ELISA (<http://cfar.ucsd.edu/core-facilities/translational-virology-core>).

■ ASSOCIATED CONTENT

Supporting Information

Quality control and assay development of the FP-based HTS, positive and negative hit selection, analysis of NC'–compound complexes, structural comparison of selected small-molecule NC' inhibitor compounds, summary of the mass spectrometry analysis of the NC'–compounds, and PubChem information on the selected compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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■ ABBREVIATIONS USED

HIV, human immunodeficiency virus; NC, nucleocapsid; NC', GST-p2-NC; p2, spacer peptide 1; Gag, group specific antigen; SL, stem-loop; Gag', GST-Gag; nt, nucleotide; rSL-2 ,

fluorescein-labeled SL-2 DNA; DSF, differential scanning fluorimetry; FP, fluorescence polarization; HTS, high-throughput screening; compd, compound; GST, glutathione S-transferase; sd, standard deviation; SEM, standard error of the mean

REFERENCES

- (1) Muriaux, D.; Darlix, J. L. Properties and functions of the nucleocapsid protein in virus assembly. *RNA Biol.* **2010**, *7*, 744–853.
- (2) Muriaux, D.; Mirro, J.; Harvin, D.; Rein, A. RNA is a structural element in retrovirus particles. *Proc. Natl. Acad. Sci. U.S.A.* **2001**, *98*, 5246–5251.
- (3) Mark-Danieli, M.; Laham, N.; Kenan-Eichler, M.; Castiel, A.; Melamed, D.; Landau, M.; Bouvier, N. M.; Evans, M. J.; Bacharach, E. Single point mutations in the zinc finger motifs of the human immunodeficiency virus type 1 nucleocapsid alter RNA binding specificities of the gag protein and enhance packaging and infectivity. *J. Virol.* **2005**, *79*, 7756–7767.
- (4) Jouvenet, N.; Simon, S. M.; Bieniasz, P. D. Imaging the interaction of HIV-1 genomes and Gag during assembly of individual viral particles. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106*, 19114–19119.
- (5) Darlix, J. L.; Garrido, J. L.; Morellet, N.; Mely, Y.; de Rocquigny, H. Properties, functions, and drug targeting of the multifunctional nucleocapsid protein of the human immunodeficiency virus. *Adv. Pharmacol.* **2007**, *55*, 299–346.
- (6) Feng, Y. X.; Campbell, S.; Harvin, D.; Ehresmann, B.; Ehresmann, C.; Rein, A. The human immunodeficiency virus type 1 Gag polyprotein has nucleic acid chaperone activity: possible role in dimerization of genomic RNA and placement of tRNA on the primer binding site. *J. Virol.* **1999**, *73*, 4251–4256.
- (7) Feng, Y. X.; Copeland, T. D.; Henderson, L. E.; Gorelick, R. J.; Bosche, W. J.; Levin, J. G.; Rein, A. HIV-1 nucleocapsid protein induces “maturation” of dimeric retroviral RNA in vitro. *Proc. Natl. Acad. Sci. U.S.A.* **1996**, *93*, 7577–7581.
- (8) Ohishi, M.; Nakano, T.; Sakuragi, S.; Shioda, T.; Sano, K.; Sakuragi, J. The relationship between HIV-1 genome RNA dimerization, virion maturation and infectivity. *Nucleic Acids Res.* **2011**, *39*, 3404–3417.
- (9) Lever, A. M.; Richardson, J. H.; Harrison, G. P. Retroviral RNA packaging. *Biochem. Soc. Trans.* **1991**, *19*, 963–966.
- (10) De Guzman, R. N.; Wu, Z. R.; Stalling, C. C.; Pappalardo, L.; Borer, P. N.; Summers, M. F. Structure of the HIV-1 nucleocapsid protein bound to the SL3 psi-RNA recognition element. *Science* **1998**, *279*, 384–388.
- (11) Amarasinghe, G. K.; De Guzman, R. N.; Turner, R. B.; Summers, M. F. NMR structure of stem-loop SL2 of the HIV-1 psi RNA packaging signal reveals a novel A-U-A base-triple platform. *J. Mol. Biol.* **2000**, *299*, 145–156.
- (12) de Rocquigny, H.; Shvadchak, V.; Avilov, S.; Dong, C. Z.; Dietrich, U.; Darlix, J. L.; Mely, Y. Targeting the viral nucleocapsid protein in anti-HIV-1 therapy. *Mini-Rev. Med. Chem.* **2008**, *8*, 24–35.
- (13) Rice, W. G.; Supko, J. G.; Malspeis, L.; Buckheit, R. W., Jr.; Clanton, D.; Bu, M.; Graham, L.; Schaeffer, C. A.; Turpin, J. A.; Domagala, J.; Gogliotti, R.; Bader, J. P.; Halliday, S. M.; Coren, L.; Sowder, R. C., 2nd; Arthur, L. O.; Henderson, L. E. Inhibitors of HIV nucleocapsid protein zinc fingers as candidates for the treatment of AIDS. *Science* **1995**, *270*, 1194–1197.
- (14) Arthur, L. O.; Bess, J. W., Jr.; Chertova, E. N.; Rossio, J. L.; Esser, M. T.; Benveniste, R. E.; Henderson, L. E.; Lifson, J. D. Chemical inactivation of retroviral infectivity by targeting nucleocapsid protein zinc fingers: a candidate SIV vaccine. *AIDS Res. Hum. Retroviruses* **1998**, *14* (Suppl. 3), S311–S319.
- (15) Chertova, E. N.; Kane, B. P.; McGrath, C.; Johnson, D. G.; Sowder, R. C., 2nd; Arthur, L. O.; Henderson, L. E. Probing the topography of HIV-1 nucleocapsid protein with the alkylating agent N-ethylmaleimide. *Biochemistry* **1998**, *37*, 17890–17897.
- (16) Druillenec, S.; Dong, C. Z.; Escaich, S.; Gresh, N.; Bousseau, A.; Roques, B. P.; Fournie-Zaluski, M. C. A mimic of HIV-1 nucleocapsid protein impairs reverse transcription and displays antiviral activity. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 4886–4891.
- (17) Druillenec, S.; Meudal, H.; Roques, B. P.; Fournie-Zaluski, M. C. Nucleomimetic strategy for the inhibition of HIV-1 nucleocapsid protein NCp7 activities. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 627–632.
- (18) Raja, C.; Ferner, J.; Dietrich, U.; Avilov, S.; Ficheux, D.; Darlix, J. L.; de Rocquigny, H.; Schwalbe, H.; Mely, Y. A tryptophan-rich hexapeptide inhibits nucleic acid destabilization chaperoned by the HIV-1 nucleocapsid protein. *Biochemistry* **2006**, *45*, 9254–9265.
- (19) Dietz, J.; Koch, J.; Kaur, A.; Raja, C.; Stein, S.; Grez, M.; Pustowka, A.; Mensch, S.; Ferner, J.; Moller, L.; Bannert, N.; Tampe, R.; Divita, G.; Mely, Y.; Schwalbe, H.; Dietrich, U. Inhibition of HIV-1 by a peptide ligand of the genomic RNA packaging signal Psi. *ChemMedChem* **2008**, *3*, 749–755.
- (20) Cruceanu, M.; Stephen, A. G.; Beuning, P. J.; Gorelick, R. J.; Fisher, R. J.; Williams, M. C. Single DNA molecule stretching measures the activity of chemicals that target the HIV-1 nucleocapsid protein. *Anal. Biochem.* **2006**, *358*, 159–170.
- (21) Stephen, A. G.; Worthy, K. M.; Towler, E.; Mikovits, J. A.; Sei, S.; Roberts, P.; Yang, Q. E.; Akee, R. K.; Klausmeyer, P.; McCloud, T. G.; Henderson, L.; Rein, A.; Covell, D. G.; Currens, M.; Shoemaker, R. H.; Fisher, R. J. Identification of HIV-1 nucleocapsid protein: nucleic acid antagonists with cellular anti-HIV activity. *Biochem. Biophys. Res. Commun.* **2002**, *296*, 1228–1237.
- (22) Warui, D. M.; Baranger, A. M. Identification of specific small molecule ligands for stem loop 3 ribonucleic acid of the packaging signal Psi of human immunodeficiency virus-1. *J. Med. Chem.* **2009**, *52*, 5462–5473.
- (23) Shvadchak, V.; Sanglier, S.; Rocle, S.; Villa, P.; Haiech, J.; Hibert, M.; Van Dorsselaer, A.; Mely, Y.; de Rocquigny, H. Identification by high throughput screening of small compounds inhibiting the nucleic acid destabilization activity of the HIV-1 nucleocapsid protein. *Biochimie* **2009**, *91*, 916–923.
- (24) Jenkins, L. M.; Byrd, J. C.; Hara, T.; Srivastava, P.; Mazur, S. J.; Stahl, S. J.; Inman, J. K.; Appella, E.; Omichinski, J. G.; Legault, P. Studies on the mechanism of inactivation of the HIV-1 nucleocapsid protein NCp7 with 2-mercaptobenzamide thioesters. *J. Med. Chem.* **2005**, *48*, 2847–2858.
- (25) Miller Jenkins, L. M.; Ott, D. E.; Hayashi, R.; Coren, L. V.; Wang, D.; Xu, Q.; Schito, M. L.; Inman, J. K.; Appella, D. H.; Appella, E. Small-molecule inactivation of HIV-1 NCp7 by repetitive intracellular acyl transfer. *Nat. Chem. Biol.* **2010**, *6*, 887–889.
- (26) Miller Jenkins, L. M.; Hara, T.; Durell, S. R.; Hayashi, R.; Inman, J. K.; Piquemal, J. P.; Gresh, N.; Appella, E. Specificity of acyl transfer from 2-mercaptobenzamide thioesters to the HIV-1 nucleocapsid protein. *J. Am. Chem. Soc.* **2007**, *129*, 11067–11078.
- (27) Amarasinghe, G. K.; De Guzman, R. N.; Turner, R. B.; Chancellor, K. J.; Wu, Z. R.; Summers, M. F. NMR structure of the HIV-1 nucleocapsid protein bound to stem-loop SL2 of the psi-RNA packaging signal. Implications for genome recognition. *J. Mol. Biol.* **2000**, *301*, 491–511.
- (28) Hill, M. K.; Shehu-Xhilaga, M.; Crowe, S. M.; Mak, J. Proline residues within spacer peptide p1 are important for human immunodeficiency virus type 1 infectivity, protein processing, and genomic RNA dimer stability. *J. Virol.* **2002**, *76*, 11245–11253.
- (29) Liang, C.; Hu, J.; Russell, R. S.; Roldan, A.; Kleiman, L.; Wainberg, M. A. Characterization of a putative alpha-helix across the capsid-SP1 boundary that is critical for the multimerization of human immunodeficiency virus type 1 gag. *J. Virol.* **2002**, *76*, 11729–11737.
- (30) Roldan, A.; Russell, R. S.; Marchand, B.; Gotte, M.; Liang, C.; Wainberg, M. A. In vitro identification and characterization of an early complex linking HIV-1 genomic RNA recognition and Pr55Gag multimerization. *J. Biol. Chem.* **2004**, *279*, 39886–39894.
- (31) Zhang, J. H.; Chung, T. D.; Oldenburg, K. R. A simple statistical parameter for use in evaluation and validation of high throughput screening assays. *J. Biol. Screening* **1999**, *4*, 67–73.
- (32) Li, F.; Goila-Gaur, R.; Salzwedel, K.; Kilgore, N. R.; Reddick, M.; Matallana, C.; Castillo, A.; Zoumplis, D.; Martin, D. E.; Orenstein, J. M.; Allaway, G. P.; Freed, E. O.; Wild, C. T. PA-457: a potent HIV

inhibitor that disrupts core condensation by targeting a late step in Gag processing. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 13555–13560.

(33) Turconi, S.; Shea, K.; Ashman, S.; Fantom, K.; Earnshaw, D. L.; Bingham, R. P.; Haupt, U. M.; Brown, M. J.; Pope, A. J. Real experiences of uHTS: a prototypic 1536-well fluorescence anisotropy-based uHTS screen and application of well-level quality control procedures. *J. Biomol. Screening* **2001**, *6*, 275–290.

(34) McGovern, S. L.; Caselli, E.; Grigorieff, N.; Shoichet, B. K. A common mechanism underlying promiscuous inhibitors from virtual and high-throughput screening. *J. Med. Chem.* **2002**, *45*, 1712–1722.

(35) Kenakin, T. *Pharmacological Analysis of Drug–Receptor Interaction*; Lippincott-Raven: Philadelphia, PA, 1997.

(36) Chan, L. L.; Pineda, M.; Heeres, J. T.; Hergenrother, P. J.; Cunningham, B. T. A general method for discovering inhibitors of protein–DNA interactions using photonic crystal biosensors. *ACS Chem. Biol.* **2008**, *3*, 437–448.

(37) Turpin, J. A. The next generation of HIV/AIDS drugs: novel and developmental antiHIV drugs and targets. *Expert. Rev. Anti-Infect. Ther.* **2003**, *1*, 97–128.

(38) Kaplan, A. H.; n, J. A.; Knigge, M.; Paul, D. A.; Kempf, D. J.; Norbeck, D. W.; Swanstrom, R. Partial inhibition of the human immunodeficiency virus type 1 protease results in aberrant virus assembly and the formation of noninfectious particles. *J. Virol.* **1993**, *67*, 4050–4055.

(39) Breuer, S.; Sepulveda, H.; Chen, Y.; Trotter, J.; Torbett, B. E. A cleavage enzyme-cytometric bead array provides biochemical profiling of resistance mutations in HIV-1 Gag and protease. *Biochemistry* **2011**, *50*, 4371–4381.

(40) Baell, J. B.; Holloway, G. A. New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays. *J. Med. Chem.* **2010**, *53*, 2719–2740.

(41) Bolton, E. E.; Wang, Y.; Thiessen, P. A.; Bryant, S. H. PubChem: integrated platform of small molecules and biological activities. *Annu. Reports Comput. Chem.* **2008**, *4*, 217–241.

(42) Vedadi, M.; Niesen, F. H.; Allali-Hassani, A.; Fedorov, O. Y.; Finerty, P. J., Jr.; Wasney, G. A.; Yeung, R.; Arrowsmith, C.; Ball, L. J.; Berglund, H.; Hui, R.; Marsden, B. D.; Nordlund, P.; Sundstrom, M.; Weigelt, J.; Edwards, A. M. Chemical screening methods to identify ligands that promote protein stability, protein crystallization, and structure determination. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 15835–15840.

(43) O'Brien, J.; Wilson, I.; Orton, T.; Pognan, F. Investigation of the Alamar Blue (resazurin) fluorescent dye for the assessment of mammalian cell cytotoxicity. *Eur. J. Biochem.* **2000**, *267*, 5421–5426.

(44) Nociari, M. M.; Shalev, A.; Benias, P.; Russo, C. A novel one-step, highly sensitive fluorometric assay to evaluate cell-mediated cytotoxicity. *J. Immunol. Methods* **1998**, *213*, 157–167.